

# Micro SpinColumns™

(25 to 75 µl Sample Volume)



a brand of Harvard Bioscience, Inc.

## Quick Start Guide

Micro SpinColumns provide rapid purification of small samples. Centrifugation or filtration under vacuum pressure can be used to run the sample through the columns. Alternatively, Micro SpinColumns may be used as Micro TipColumns™ by passing the sample through the column using a micro-pipette. Each package includes a column, two 2 ml centrifuge tubes, and top cap (for gel filtration) or frit. Available with our complete range of packing materials or pre-packed with custom requested materials.

### Instructions for use as SpinColumns

1. Place the column into a centrifuge tube. For gel filtration media, tap the column gently to ensure that the media is settled at the bottom and remove the blue cap.
2. Place 150 µl of water or buffer in the column and wait 10 minutes for hydration.
3. Centrifuge for 2 to 3 minutes at approximately 1,000 x g.
4. Repeat Steps 2 and 3 if desired.
5. Remove column from tube and blot the exterior dry.
6. Add between 25 µl and 75 µl of sample to the column.
7. Place the column in a new centrifuge tube and spin for 2 to 3 minutes at approximately 1,000 x g.

#### For size exclusion applications:

- a) The purified sample is collected in the centrifuge tube.

#### For solid-phase extraction technique:

- a) Unbound sample components are removed.
- b) Place column into a new centrifuge tube, add elution buffer, and centrifuge to recover desired sample.



### Instructions for use as TipColumns

1. For Gel Filtration media, tap the column gently to ensure that the media is settled at the bottom and remove the blue cap.
2. Place 150 µl of water or buffer in the column and wait 10 minutes for hydration.
3. Dispense excess liquid.
4. Add between 25 µl and 75 µl of sample to the column.

#### For size exclusion applications:

- a) Add sample to top of the tip and pressure into the media.
- b) Add elution buffer to top of tip and collect into sample tube.
- c) Optionally, steps a) to b) may be repeated 2x as necessary.

#### For solid phase extraction technique:

- a) Aspirate sample into tip or add to top of tip.
- b) Dispense unbound sample.
- c) Repeat steps a) and b) as necessary to further remove unbound sample components.
- d) Add elution buffer and collect purified sample.

*Micro SpinColumns are intended for single use only.*

## Ordering Information

Empty SpinColumns		
Frit	Qty. of 24	Qty. of 96
5 to 10 µm frit	74-4421	74-4420
20 µm frit	74-4401	74-4400
40 µm frit	74-4431	74-4430
Filled SpinColumns		
Media Type	Qty. of 24	Qty. of 96
Ion Exchange		
Strong Anion Q	74-4704	74-4700
Weak Anion PEI	74-4411	74-4410
Weak Anion DEAE	74-4705	74-4701
Strong Cation SA	74-4413	74-4412
Strong Cation SP	74-4706	74-4702
Weak Cation CM	74-4707	74-4703
Weak Cation AA	74-4415	74-4414
Gel Filtration		
Sephadex, G-10 (700 D)	74-4504	74-4500
Sephadex, G-25 (5 kD)	74-4505	74-4501
Sephadex, G-50 (30 kD)	74-4506	74-4502
Sephadex, G-100 (100 kD)	74-4507	74-4503
Polyacrylamide, P-2 (2 kD)	74-4808	74-4802
Polyacrylamide, P-6 (6 kD)	74-4809	74-4803
Hydrophilic (Normal Phase)		
Amino (NH <sub>2</sub> )	74-4611	74-4605
Cyano (CN)	74-4610	74-4604
PHEA	74-4811	74-4805
Silica	74-4606	74-4600
Hydrophobic (Reverse Phase)		
C4	74-4609	74-4603
C8	74-4608	74-4602
C18	74-4607	74-4601
C18 Targa	74-4613	74-4614
Misc.		
Activated Charcoal	74-4806	74-4800
Cellulose	74-4807	74-4801
Detergent Removal	74-4810	74-4804

Key:

Q = Quaternary Ammonium (Sephacrose, Fast Flow)  
 PEI = Linear Polyethyleneimine (Silica Based: Organic Compatible)  
 DEAE = Cross-Linked Diethylaminoethyl (Sephacrose)  
 PHEA = Hydrophilic Polyhydroxyethyl Aspartamide

SA = Sulfoethyl Aspartamide (Silica Based: Organic Compatible)  
 CM = Carboxymethyl 12 µm, 300 Å (Sephacrose)  
 SP = Sulfopropyl (Sephacrose, Fast Flow)  
 AA = Aspartic Acid 20 µm, 300 Å (Silica Based: Organic Compatible)